



TRAINING COURSE ON

THE ANALYSIS OF FOOD AND FEED SAMPLES FOR THE PRESENCE OF GENETICALLY MODIFIED ORGANISMS

26 - 30 October, 2009

Centro de Ingeniería Genética y Biotecnología (CIGB) La Habana, Cuba



COURSE CONTENT

Covered topics:

- Overview of EU legislation on GMOs and specific requirements
- Introduction on the general procedures for GMO detection
- Experimental planning and sample preparation
- DNA extraction
- Qualitative PCR for GMO analysis
- □ Real-time PCR for GMO quantification
- Sampling concepts and recommended EU protocol
- Laboratory implementation and conduction of a GMO detection laboratory
- □ Implementation of ISO 17025 for accreditation in a GMO testing laboratory
- Method validation criteria

Experimental work:

- Sample preparation and DNA extraction
- Qualitative PCR
- □ GMO quantitative analysis by real-time PCR
- Data analysis, expression and interpretation of the results

The course will provide overall scientific and technical information on sampling and on the analytical approaches for GMO analysis as well as hands on experience on how the methods are performed in the laboratory. In addition, the course will prove information on the different theoretical and technical requirements for proper laboratory implementation and conduction of testing activity according to current legislative requirements.

WORKPROGRAMME

1ST DAY – MONDAY, OCTOBER 26TH 2009

9:00	am	Welcoming speech – opening of the course – C. Borroto - CIGB
9:30	am	Introduction and course content, presentation of the organizers and of the
		participants – M. Querci - JRC

PREPARATION OF SAMPLES: DNA EXTRACTION

10:10	am	Experimental: DNA extraction following the CTAB method - Part 1
10:40	am	Coffee break
11:00	am	Experimental: Preparation of agarose gels DNA extraction following the CTAB method - Part 2
1:00	pm	Lunch
2:00	pm	Experimental: DNA extraction following the CTAB method - Part 3
3:00	pm	Experimental: Sample loading
3:45	pm	Coffee break
4:00	pm	Theory: Sample preparation and DNA extraction
5:00	pm	Experimental: Interpretation of the gels
5:20	pm	End of the day

2ND DAY – TUESDAY, OCTOBER 27TH 2009

QUALITATIVE PCR

9:00	am	Theory: Overview of GMO testing methodology
10:00	am	Experimental: Qualitative PCR Plant specific: detection of the zein and lectin genes
10:45	am	Coffee break
11:00	am	Preparation of agarose gels
11:20	am	Theory: The polymerase chain reaction (PCR) and its application in GMO analysis
12:20	pm	Lunch
1:30	pm	Sample loading
2:00	pm	<u>Theory:</u> GMO testing laboratory implementation, quality system and quality assurance
3:00	pm	Interpretation of the gels (zein and lectin specific PCR)
3:15	pm	Coffee break
3:45	pm	Experimental: Screening PCR: detection of the 35S promoter
4:45	pm	End of the day

3RD DAY – WEDNESDAY, OCTOBER 28TH 2009

QUALITATIVE PCR

9:00	am	Experimental: Nested PCR for the specific detection of Roundup Ready® soybean (1st PCR reaction)
10:00	am	Preparation of agarose gels
10:30	am	Coffee break
11:00	am	<u>Theory</u> : Serological and novel methodological approaches for the detection of GMOs
12:00	pm	Sample loading (35S screening PCR)
12:30	pm	Lunch
1:30	pm	Experimental: Nested PCR for the specific detection of Roundup Ready® soybean (2 nd PCR reaction)
2:30	pm	Experimental : Preparation of agarose gels and interpretation of the 35S screening PCR
3:00	pm	Coffee break
3:30	pm	Theory: Real-time PCR and its application in GMO analysis: detection and quantification
4:45	pm	Experimental: Sample loading (nested PCR products)
5:15	pm	End of the day

4TH DAY – THURSDAY, OCTOBER 29TH 2009

QUANTITATIVE REAL-TIME PCR

9:00	am	Experimental : Interpretation of the gel Roundup Ready soybean specific nested PCR
9:15	am	Experimental: Preparation of samples for the Real-Time PCR and samples loading
10:30	am	Coffee break
11:00	am	Theory: Real-time PCR and its application in GMO analysis – Part 2
12:15	pm	Lunch
1:15	pm	Experimental: Experimental design, data analysis and interpretation
2:45	pm	<u>Theory:</u> EU legislation on GMOs and mandate of the European Commission Joint Research Centre (JRC)
3:45	pm	Coffee break
4:00	pm	Theory: Sampling: basic principles
5:00	pm	End of the day

5TH DAY – FRIDAY, OCTOBER 30TH 2009

QUANTITATIVE REAL-TIME PCR

9:00 am	Experimental: Preparation of samples for the Real-Time PCR and samples loading
10:00 am	Coffee break
10:30 am	<u>Theory:</u> Method validation/verification and introduction to measurement uncertainty
12:00 pm	Lunch
1:30 pm	Experimental: Data analysis, result interpretation and reporting
2:30 pm	Round table: Troubleshooting, data interpretation and practical experimental issues. Questions and answers session. General discussion and conclusion of the course
4:00 pm	Transport to the hotel or airport